



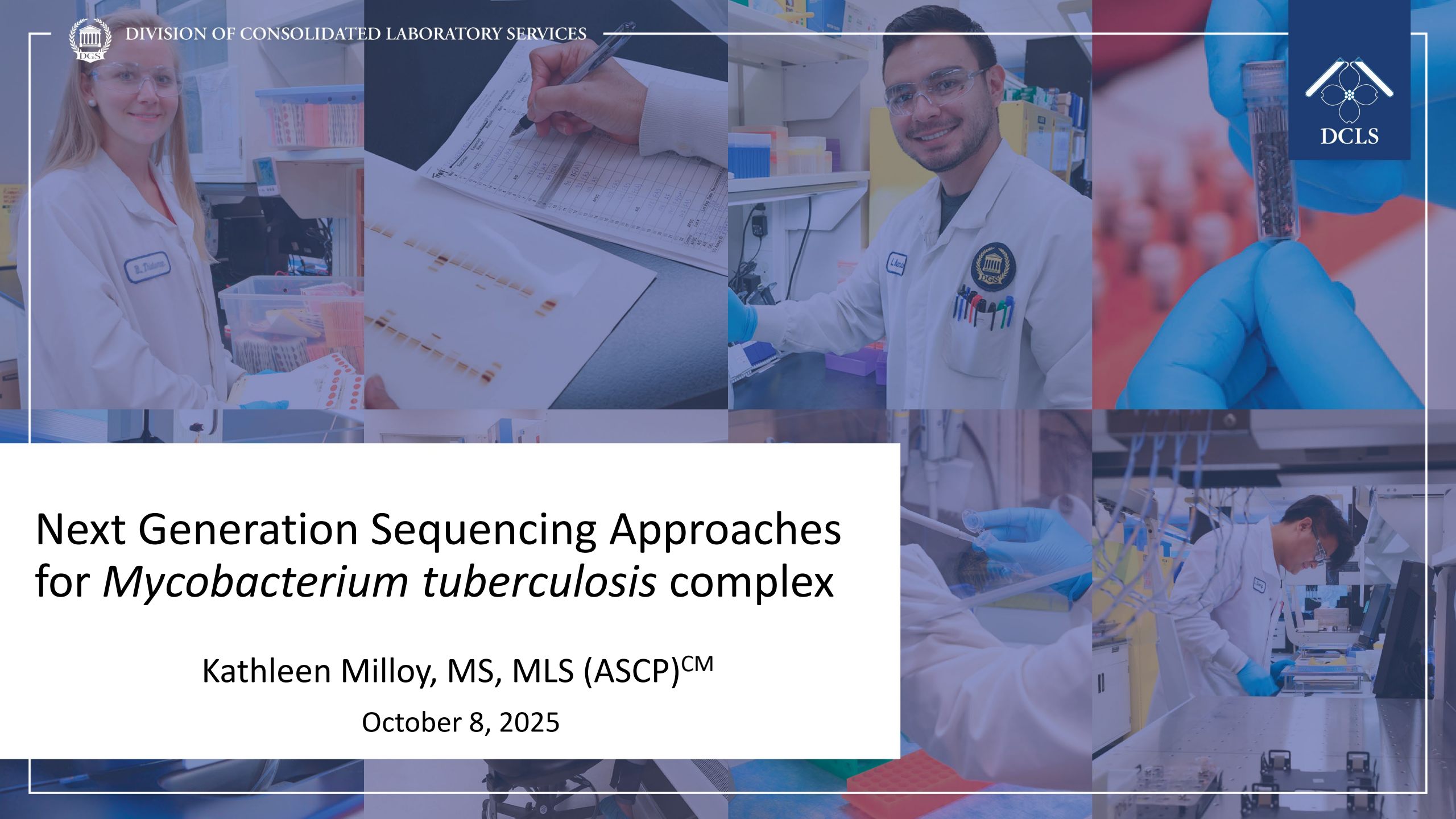
DIVISION OF CONSOLIDATED LABORATORY SERVICES



# Next Generation Sequencing Approaches for *Mycobacterium tuberculosis* complex

Kathleen Milloy, MS, MLS (ASCP)<sup>CM</sup>

October 8, 2025



# Objectives

- Explain the utility of next generation sequencing (NGS) and the most common methods used for *Mycobacterium tuberculosis* complex (MTBC)
- Describe a NGS workflow for MTBC
- Provide an overview of drug resistant MTBC and susceptibility testing
- Discuss the WHO catalog of mutations
- Describe how targeted next generation sequencing (tNGS) is utilized to detect resistance markers in MTBC
- Provide information for Reference Laboratory susceptibility testing of MTBC
  - CDC MDDR service and Susceptibility Testing Reference Center
  - CA MDL National DST Reference Lab

# Next Generation Sequencing (NGS)

- Large-scale, high-throughput DNA sequencing
- Generates a large amount of data that can be used for a variety of purposes

Detection of mutations  
that predict drug  
resistance and  
susceptibility

Identification of strains  
or species

Compare genotypes

High quality analysis to  
determine relatedness  
to other cases

# Polling Question 1

**What is the definition of a gene?**

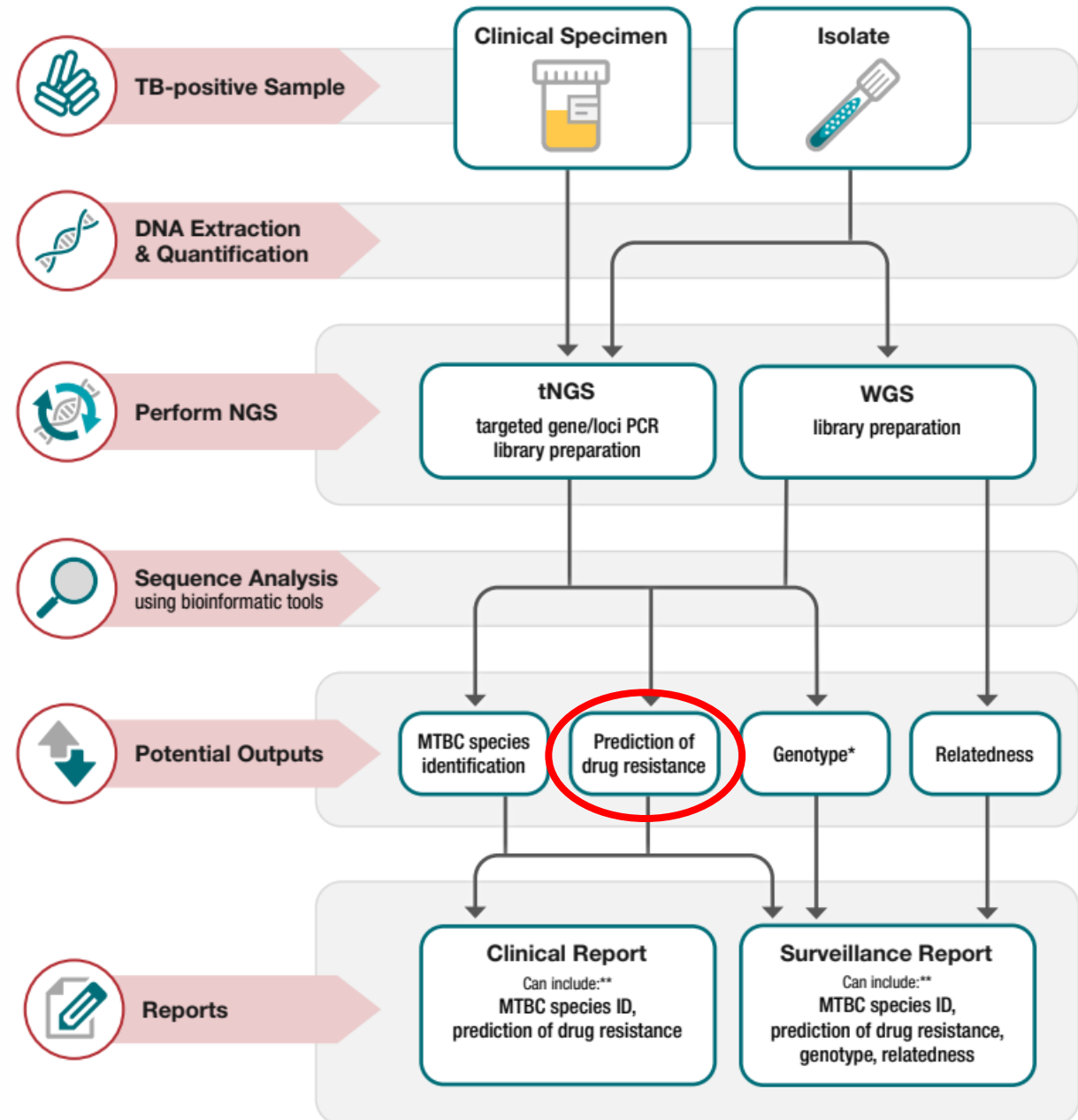
- a) A specific sequence of DNA that contains the information for making a protein
- b) An enzyme responsible for making nucleotides
- c) A type of cell organelle that contains DNA
- d) A random sequence of nucleotides in a chromosome



# NGS Methods

Targeted NGS (tNGS)	Whole genome sequencing (WGS)
Amplicon-based NGS, to sequence specific parts of the genome (i.e. single gene or group of genes)	Sequence the entire genome of an organism
Useful for diagnostic purposes	Can be used for diagnostic and epidemiological/surveillance purposes
Can be used on primary specimens and isolates	Can be used on isolates

# TB NGS Workflow

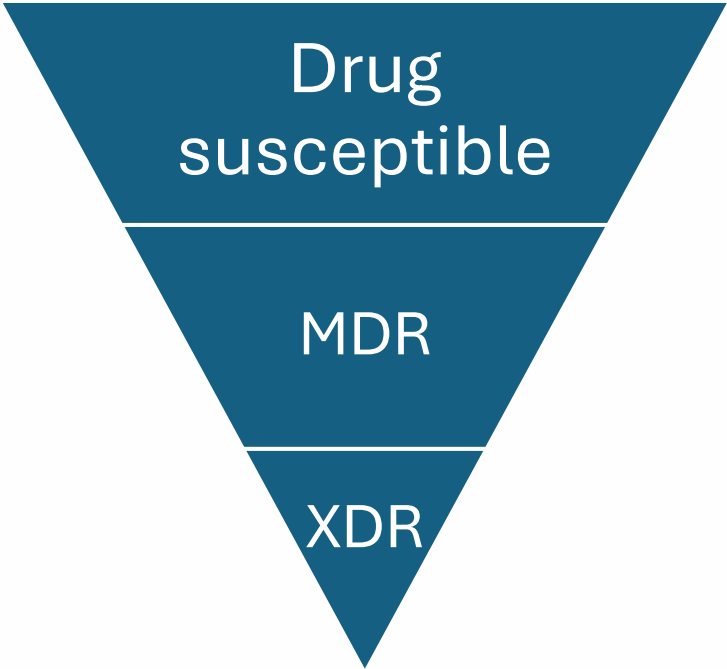


Reference: [APHL TB NGS Fact Sheet](#)





# Drug Resistant *Mycobacterium tuberculosis* complex (MTBC)



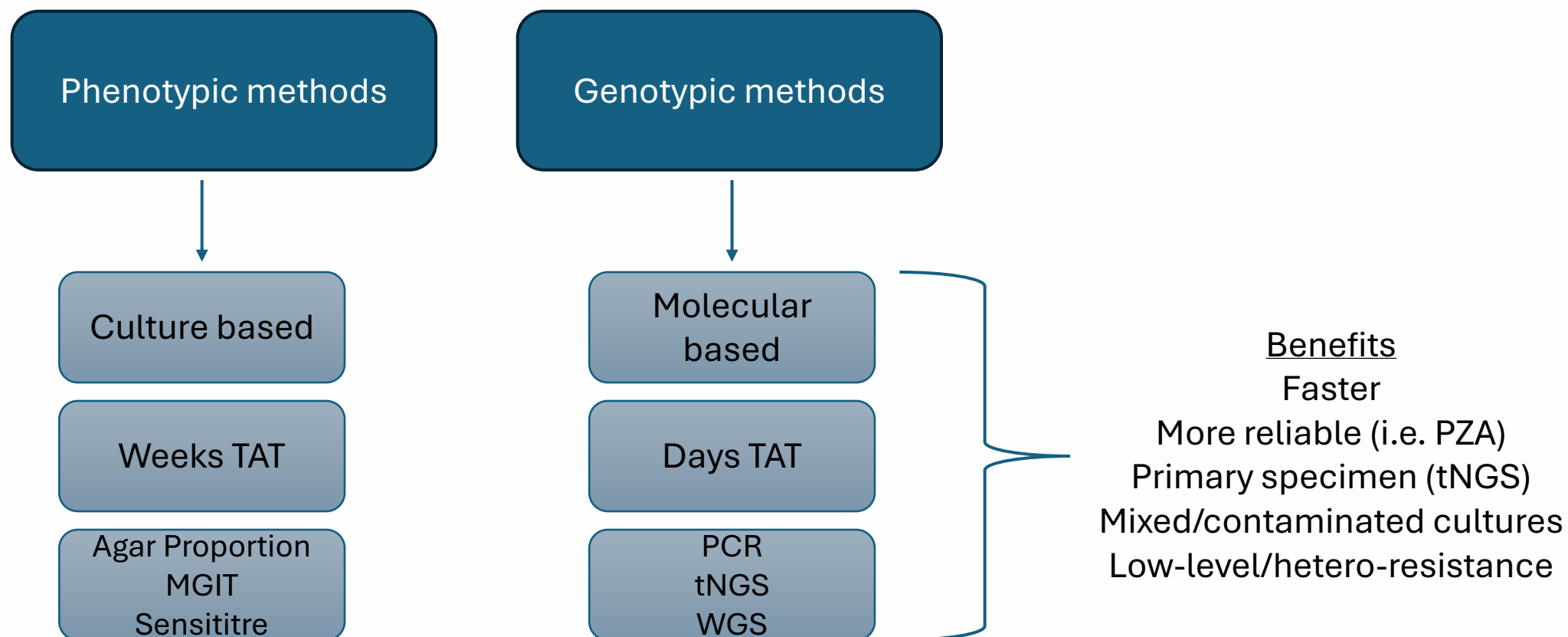
% Drug Resistance: United States (2023 CDC data)

INH Resistant	Multidrug-resistant (MDR)	Pre-extensively drug-resistant (Pre-XDR)
8.5%	1.4%	0.2%

% Drug Resistance Among TB cases: Virginia (2024 VDH data)

First-line Resistant	Multidrug-resistant (MDR)
9.8%	0%

# Drug Susceptibility Testing (DST)





# Genetic Terminology

## Variant or Mutation

Alteration in the nucleotide sequence of an organism that may or may not have a phenotypic effect

## Silent Mutation

Change to a nucleotide sequence that does not result in a change to the amino acid

## Single Nucleotide Polymorphism

A variation at a single nucleotide position; the most common type of mutation causing drug resistance in MTB

## Wild Type (WT)

Standard genetic sequence to which mutations are compared (H37Rv commonly used WT (pan-susceptible) strain for MTB)

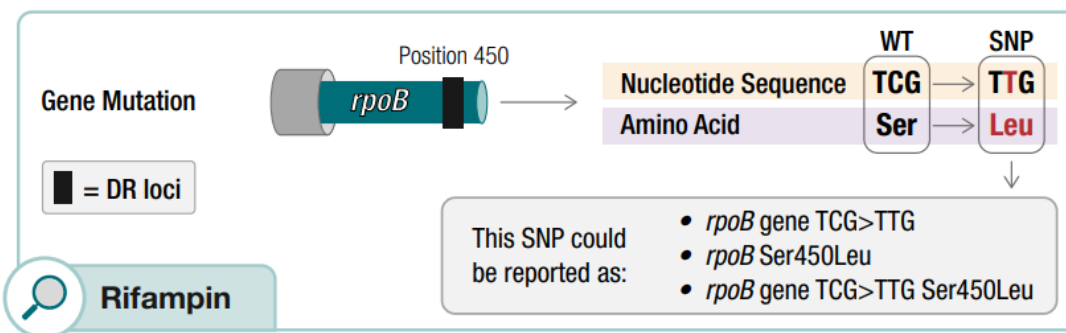
# Molecular DST (mDST) for MTBC

- As NGS capabilities expand, laboratories are using mDST to predict MTBC drug resistance
- Laboratory availability varies based on many factors:
  - Incidence rate
  - Infrastructure
  - Expertise
  - Laboratory developed tests (LDTs)

Reference: [APHL NGS and Mutations](#)

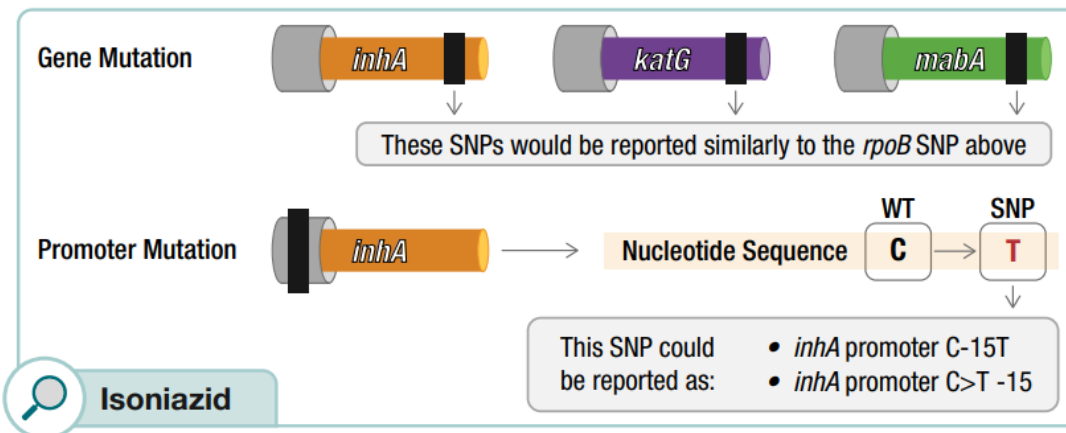
## Mutations that confer DR can be simple...

Sometimes resistance to a drug is associated with a single gene or area of a gene.



## ...or complicated.

Resistance may stem from a mutation within one of several genes or their promoters, making identification more challenging.



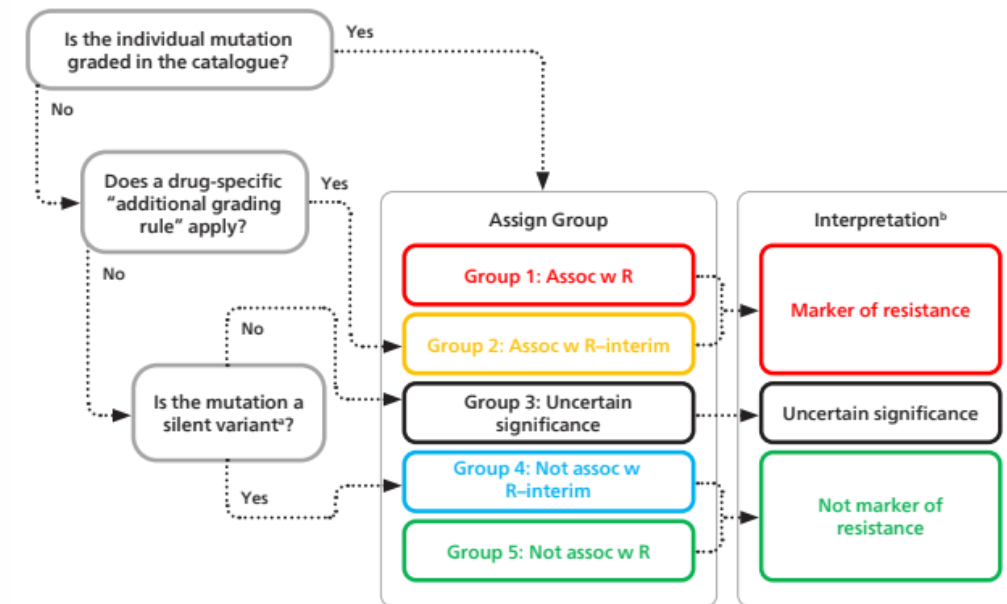
# Catalogue of mutations in *Mycobacterium tuberculosis* complex and their association with drug resistance

Second edition



- Large database of WGS and phenotypic drug susceptibility testing data from analysis of global clinical isolates
- WHO recommends routine testing of all TB patients for rapid detection of resistance to rifampin and isoniazid

Fig. 1. Instructions for using the catalogue



DCLS

**Table 3. Groups 1–5 mutations for the second edition and their performance for predicting phenotypic resistance**

		Group 1: Assoc w R	Group 2: Assoc w R-interim	Group 3: Uncertain significance	Group 4: Not assoc w R-interim	Group 5: Not assoc w R
<b>RIF</b>	No. of variants identified	26	110	4484	2 (2568)	52 (32)
	Sens, spec, PPV (% [95% CI])	92.1 (91.7–92.5), 97.1 (96.9–97.3), 94.5 (94.2–94.9)	1.1 (1.0–1.3), 99.8 (99.8–99.9), 76.5 (70.8–81.6)			
	<b>Combined performance</b>	<b>93.3 (92.9–93.7), 96.9 (96.7–97.1), 94.2 (93.9–94.6)</b>				
<b>INH</b>	No. of variants identified	7	135	5404	11 (1671)	41 (16)
	Sens, spec, PPV (% [95% CI])	89.6 (89.2–90.0), 98.2 (98.1–98.4), 97.5 (97.2–97.7)	2.0 (1.8–2.2), 99.7 (99.6–99.7), 82.5 (78.9–85.7)			
	<b>Combined performance</b>	<b>91.6 (91.2–92.0), 97.9 (97.8–98.1), 97.1 (96.8–97.3)</b>				
<b>EMB</b>	No. of variants identified	13	0	4943	10 (2068)	50 (34)
	Sens, spec, PPV (% [95% CI])	81.1 (80.3–81.9), 91.6 (91.3–91.9), 71.9 (71.0–72.8)	0 (0–0), 100.0 (100.0–100.0), 0 (0–0)			
	<b>Combined performance</b>	<b>81.1 (80.3–81.9), 91.6 (91.3–91.9), 71.9 (71.0–72.8)</b>				
<b>PZA</b>	No. of variants identified	139	202	1465	20 (720)	17 (8)
	Sens, spec, PPV (% [95% CI])	63.5 (62.0–64.9), 98.6 (98.5–98.8), 92.4 (91.4–93.4)	14.6 (13.5–15.6), 99.2 (99.1–99.3), 82.9 (80.1–85.5)			
	<b>Combined performance</b>	<b>78.0 (76.8–79.2), 97.9 (97.6–98.1), 90.5 (89.5–91.4)</b>				
<b>LFX</b>	No. of variants identified	12	6	2016	2 (983)	19 (9)
	Sens, spec, PPV (% [95% CI])	83.6 (82.6–84.5), 97.3 (97.0–97.5), 89.2 (88.4–90.0)	1.2 (1.0–1.6), 99.6 (99.6–99.7), 48.3 (40.1–56.6)			
	<b>Combined performance</b>	<b>84.8 (83.9–85.7), 96.9 (96.7–97.1), 88.1 (87.3–89.0)</b>				

- Resistant
- Likely resistant

- Unknown
- Undetermined

- Susceptible
- Sensitive
- Likely susceptible

# CDC Test Offering

CDC TB Laboratory	CDC Pathology
Molecular Detection of Drug-Resistance Service (MDDR)	Infectious disease testing for human tissues (when culture is not available)
Phenotypic DST (agar proportion, first and second-line)	Histopathology, histochemistry, immunohistochemistry, PCR and sequencing
Samples must be routed through state health department/laboratory or authorized facility Pre-approval required for testing	



CSTOR

Specimen Test Order and Reporting

Welcome, Kathleen Milloy

Division of Consolidated Laboratory Services



Create Test Order Requests

Submit Specimens

Ship Package

Check Status

View Reports

Training

My Needs Attention - 0

You do not have any notifications.

Organization Status

7 Days

Test Orders  
Submitted



3

Draft  
Specimens



0

Specimens In  
Transit



0

Specimens  
Received



0

Packages  
Ready For  
Shipment



0

Reports  
Released



5



DIVISION OF CONSOLIDATED LABORATORY SERVICES

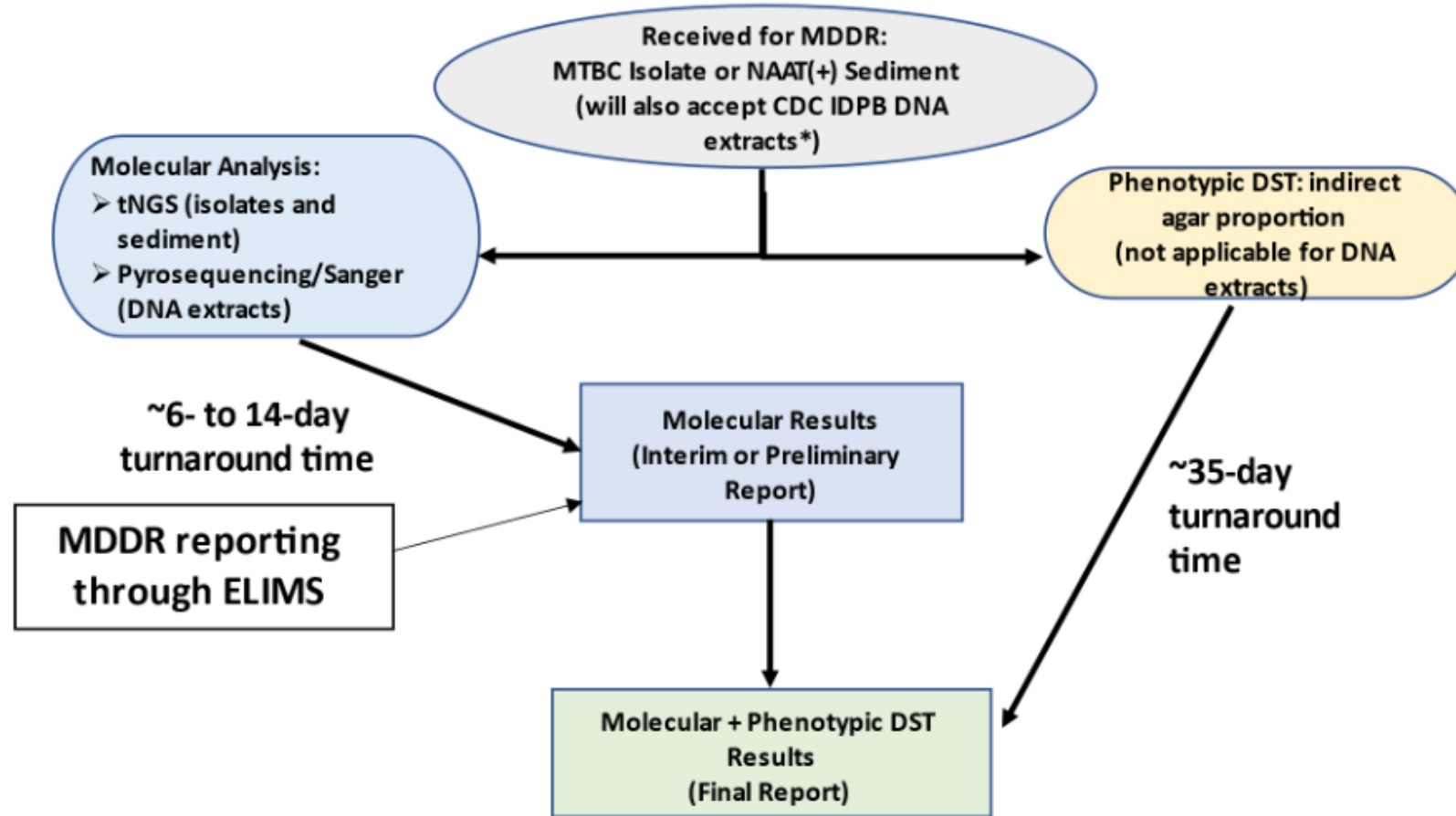


# MDDR Testing at CDC

- Utilizes DNA sequencing for detection of mutations most frequently associated with resistance to first and second-line drugs as well as new/repurposed drugs such as bedaquiline, clofazimine, and linezolid
- tNGS assay that is CLIA compliant
  - DNA regions of interest are amplified by PCR
  - Sequencing libraries generated using Illumina Nextera XT Library Prep Kit
  - Libraries sequenced on Illumina MiniSeq platform
- All isolates (including those grown from sediments) will also undergo growth-based DST
- [MDDR User Guide](#)



# MDDR Algorithm



\*DNA extracts only accepted from CDC IDPB and will be tested by conventional sequencing methods (not tNGS)

MTBC *Mycobacterium tuberculosis* complex, IDPB: Infectious Diseases Pathology Branch, ELIMS: enterprise laboratory information management system

Reference: [MDDR User Guide](#)



# Genetic Loci Examined in MDDR Assay

Drug	Genetic locus
Rifampin	<i>rpoB</i>
Isoniazid	<i>katG, fabG1- inhA</i>
Ethambutol	<i>embB</i>
Pyrazinamide	<i>pncA</i>
Fluoroquinolones	<i>gyrA, gyrB</i>
Amikacin Capreomycin Kanamycin	<i>rrs</i>
Kanamycin	<i>eis</i>
Bedaquiline	<i>AtpE, rv0678, pepQ</i>
Clofazimine	<i>Rv0678, pepQ</i>
Linezolid	<i>RplC, rrl</i>



Assistance with MDDR results interpretation:

Laboratorians, TB control personnel, and healthcare providers can contact the CDC by email ([TBLab@cdc.gov](mailto:TBLab@cdc.gov)) or telephone (404-639-2455) for help in interpretation of reports



Centers for Disease Control and Prevention  
National Tuberculosis Reference Laboratory

Patient Name: [REDACTED]  
Sex: [REDACTED] Birthdate: [REDACTED] Age: [REDACTED] Date of Onset: [REDACTED]

Public Health / International Submitter IDs

Patient ID: [REDACTED] Alt. Patient ID: [REDACTED]  
Specimen ID: [REDACTED] Alt. Specimen ID: [REDACTED]

CDC Specimen ID: [REDACTED] CDC Unique ID: [REDACTED] CDC Local Aliquot ID: [REDACTED]

**Rifampin (RIF)** **Result** **Interpretation**

RIF interpretation RIF resistant  
rpoB\* Ser450Leu

Comments and Disclaimers

\* DTBE Reference Laboratory has transitioned from the E. coli to the M. tuberculosis numbering system for reporting rpoB gene mutations.

**Isoniazid (INH)** **Result** **Interpretation**

INH interpretation INH resistant  
inhA No mutation  
fabG1 No mutation  
katG Partial amplification,  
Ser315Thr

**Ethambutol (EMB)** **Result** **Interpretation**

EMB interpretation Likely EMB resistant  
embB Met306Ile

**Pyrazinamide (PZA)** **Result** **Interpretation**

PZA interpretation Likely PZA resistant  
pncA Ile90Thr

**Fluoroquinolones (FQ)** **Result** **Interpretation**

FQ interpretation Cannot rule out FQ resistance.  
gyrA No mutation  
gyrB No mutation



Centers for Disease Control and Prevention  
National Tuberculosis Reference Laboratory

Patient Name: [REDACTED]  
Sex: Female Birthdate: [REDACTED] Age: [REDACTED] Date of Onset: [REDACTED]

Public Health / International Submitter IDs

Patient ID: [REDACTED] Alt. Patient ID: [REDACTED]  
Specimen ID: [REDACTED] Alt. Specimen ID: [REDACTED]

CDC Specimen ID: [REDACTED] CDC Unique ID: [REDACTED] CDC Local Aliquot ID: [REDACTED]

**MTBC Agar Proportion Susceptibility\*** **% Resistant** **Interpretation**

Isoniazid 0.2 µg/mL	100 %	Resistant
Isoniazid 1.0 µg/mL	100 %	Resistant
Isoniazid 5.0 µg/mL	100 %	Resistant
Rifampin 1.0 µg/mL	100 %	Resistant
Ethambutol 5.0 µg/mL	50 %	Resistant
Streptomycin 2.0 µg/mL	100 %	Resistant
Streptomycin 10.0 µg/mL	50 %	Resistant
Rifabutin 2.0 µg/mL	50 %	Resistant
Ciprofloxacin 2.0 µg/mL	0 %	Susceptible
Kanamycin 5.0 µg/mL	0 %	Susceptible
Ethionamide 10.0 µg/mL	100 %	Resistant
Capreomycin 10.0 µg/mL	0 %	Susceptible
PAS 2.0 µg/mL	0 %	Susceptible
Ofloxacin 2.0 µg/mL	0 %	Susceptible
Amikacin 4.0 µg/mL	0 %	Susceptible

Comments and Disclaimers

\* Susceptibility testing method: Indirect agar proportion, 7H10 medium. Resistance is defined as >1% (growth on drug-containing medium compared to drug-free medium).  
This test has not been cleared or approved by the FDA. The performance characteristics have been established by the DTBE Reference Laboratory.

**MTBC Pyrazinamide Susceptibility\*** **Result**

Pyrazinamide 100 µg/mL<sup>†</sup> Not Tested

Comments and Disclaimers

<sup>†</sup> Test not done  
This test order is unavailable until further notice.  
\* Susceptibility testing method: Mycobacteria Growth Indicator Tube (MGIT)




### Rifamycins (RIF) & *rpoB* Gene

- Rifamycins include: rifampin (rifampicin), rifabutin and rifapentine.
- ~95% of DR mutations are found in the 81-basepair RIF Resistance Determining Region (RRDR).
- Although less common, mutations outside of the RRDR can confer DR.
- A subset of mutations confer low-level resistance and may not be detected by pDST.

 ***rpoB* Ser450Leu, His445Tyr, His445Asp, Asp435Val**


### Isoniazid (INH) & *katG* Gene

- *katG* mutations are responsible for ~85% of observed INH DR.
- Most frequent *katG* mutation is Ser315Thr, though other *katG* variants also cause resistance.
- Mutations in the promoter and genes for *inhA* and *fabG1(mabA)* may also be associated with INH DR.
- Silent *fabG1(mabA)* mutation Leu203Leu is also known to be associated with INH DR.

 ***katG* Ser315Thr**

### Ethionamide (ETH) & *inhA* and *ethA* Genes

- INH and ETH are structural analogs and cross-resistance is common.
- Mutations in *ethA* are known to be associated with ETH DR.
- *inhA* promoter mutation C-15T is often associated with low-level INH and ETH resistance.
- Mutations in *fabG1(mabA)* are also associated with ETH DR.

 ***inhA* C-15T**


### Pyrazinamide (PZA) & *pncA* Gene/Promoter

~85% of *pncA* genetic variants are associated with phenotypic PZA DR.

 **All *M. bovis* (including BCG strain) have a *pncA* His57Asp mutation and are resistant to PZA**


### Ethambutol (EMB) & *embB* Gene

- SNPs in *embA*, *embC* and the *embC-embA* promoter region have also been associated with EMB DR.
- Not all EMB DR associated mutations are known and discordance between methods can be observed.

 ***embB* Met306Val**

### Fluoroquinolone (FQ) & *gyrA* Gene

- FQs used for MTB treatment include moxifloxacin, levofloxacin and ofloxacin.
- Mutations within the quinolone-resistance-determining region (QRDR) of DNA gyrase subunit *gyrA* are most frequently linked to DR; mutations in the QRDR of *gyrB* can also be linked to DR.
- Heteroresistance is common and can cause discordant results.

 ***gyrA* Asp94Gly, Ala90Val**

### Second Line Injectable Drugs & *rrs*, *eis* and *tlyA* Genes

Second line injectable drugs include amikacin, kanamycin and capreomycin.

 ***rrs* A1401G**

### New TB Drugs

The following drugs and genes have known DR associations:

- Bedaquiline & Rv0678 and *atpE*
- Linezolid & *atpE*, *rplC* and *rml*
- Delamanid and pretomanid & *fgd1*, *ddn*, *fbiA*, *fbiB*, *fbiC* and *fbiD*
- Clofazimine & *pepQ*, Rv0678, *mmpL5* and *mmp*

Note that newer drugs have less robust DR data, so continued tracking is essential.

\*For more information, see WHO Catalogue of mutations in *Mycobacterium tuberculosis* complex and their association with drug resistance, 2nd edition.



# Polling Question 2

**Which of the following genetic loci is used for determining rifampin resistance?**

a) *pncA*

b) *katG*

c) *rpoB*

D) *inhA*



# National Public Health Laboratory DST Reference Center for *Mycobacterium tuberculosis*



- Provides DST for eligible US PHL with low volumes of testing
- Performed at California Department of Public Health's Microbial Diseases Laboratory (MDL)
  - First-line DST using MGIT (rifampin, isoniazid, ethambutol, moxifloxacin)
  - Pyrazinamide DST using MGIT when a mutation with uncertain significance is detected via molecular methods and PZA resistance cannot be ruled out
  - Second-Line DST using MGIT (ethionamide, capreomycin, amikacin, moxifloxacin, kanamycin and rifabutin)
  - Molecular detection of drug resistance performed by WGS on submitted isolates and by tNGS on specimen sediments, mixed and non-viable cultures





# MDL WGS and tNGS DST Interpretation Summary

Mutation Interpretation		Individual mutation reporting		Drug interpretation reporting
		Format of individual mutations reported per gene locus	Mutation is listed in clinical report (Yes/No)	Overall value based on the highest severity mutation within all targets associated with given drug
<b>R</b>	R-mutations for all targets, except for <i>rpoB</i> (see below)	c.2066C>T (p.Ala689Val)	Yes	Mutation(s) associated with resistance to XXX detected
	R mutations in <i>rpoB</i> gene	Low-level RIF R mutations	Yes	Predicted low-level resistance to rifampin. May test susceptible by phenotypic methods
		Other RIF R mutations	Yes	Predicted resistance to rifampin
<b>U</b>		c.2066C>T (p.Ala689Val)	Yes	The detected mutation(s) have uncertain significance. Resistance to XXX cannot be ruled out
<b>S</b>	S-mutations for all targets, except for <i>rpoB</i> (see below)	No high confidence mutations detected	No	No mutations associated with resistance to XXX detected
	S mutations in <i>rpoB</i> gene	Synonymous c.2066C>T (p.Ala689Ala) [synonymous] <sup>1</sup>	Yes	Predicted susceptibility to rifampin. The detected synonymous mutation(s) do not confer resistance <sup>2</sup> Additionally displayed in comments: The detected in <i>rpoB</i> synonymous mutation may result in false-resistance in PCR-based assays targeting the <i>rpoB</i> RRDR
		Other RIF S mutations (outside of RRDR)	No	Predicted susceptibility to rifampin <sup>2</sup>
<b>WT</b>		No mutations detected	N/A	No mutations associated with resistance to XXX detected
<b>Insufficient Coverage</b> in the gene locus <sup>3</sup> AND Successfully sequenced areas of the same gene OR other genes associated with given drug do NOT contain mutations interpreted as R		No sequence <sup>4</sup>	N/A	Not all targets could be sequenced; resistance to XXX cannot be ruled out <sup>5</sup>



Reference: [National PHL DST Reference Center Reporting Language and Bioinformatic Pipeline](#)



# National TB Molecular Surveillance Center

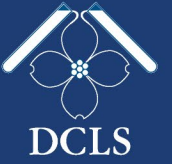


- CDC contracts with Michigan laboratory to provide genotyping services
- Provides support for epidemiologic investigations and surveillance activities
- Performs WGS to distinguish different strains of *M. tuberculosis*
  - Helps distinguish between persons whose TB disease is the result of TB infection that was acquired in the past, compared to recently or newly acquired.
  - Add value to conventional contact investigations
  - Offers a tool to help direct TB prevention and control efforts to preventing transmission
  - Allows monitoring of our progress toward eliminating TB transmission
- State PHL submits index isolate for each patient (1 per year)
- TB Genotyping Information Management System



# WGS Analysis

Type of Analysis	Purpose
Whole-genome multilocus sequence typing (wgMLST)	<ul style="list-style-type: none"> <li>Analyzes 2,672 gene/loci and assigns a number for each.</li> <li>Isolates that match at <math>\geq 99.7\%</math> of loci form a genotype cluster with a wgMLSType name</li> </ul>
Whole-genome single nucleotide polymorphism (wgSNP) comparison	<ul style="list-style-type: none"> <li>Identify SNPs that distinguish isolates in a genotype-matched cluster.</li> <li>Number of SNPs that differ between isolates can provide information about recent transmission.</li> <li>SNPs used to map phylogenetic tree</li> </ul>
Detection of possible drug resistance	<ul style="list-style-type: none"> <li>Detect mutations associated with drug resistance for surveillance purposes</li> </ul>



# Contact

[Kathleen.milloy@dgs.virginia.gov](mailto:Kathleen.milloy@dgs.virginia.gov)

(804)648-4480 ext. 223



DIVISION OF CONSOLIDATED LABORATORY SERVICES