

Overview

- Bloom response ODU
- Notable 2020 blooms
- Monitoring results summary
- Related projects –transport, IFCB
- 2021

VDH Shellfish monitoring

- Monthly collections- routine fixed sites
 - Lugol's solution (500mL) phytoplankton analyses (ODU)
 - Unpreserved frozen sample (50mL)- ELISA screening (VDH)
- Bloom samples
 - Response to bloom reports or visual observation by field staff
 - VDH, CBP, HRSD, Time series site

National Shellfish Sanitation Program (NSSP)

Guide for the Control of Molluscan Shellfish 2017 Revision

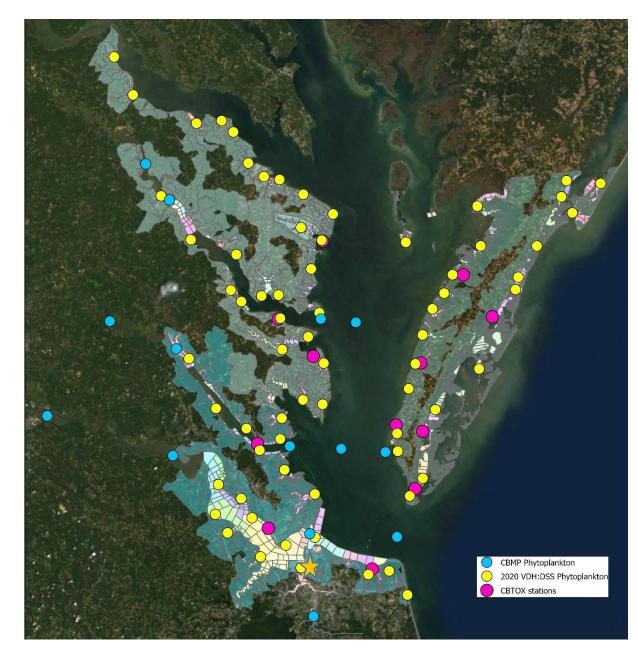


From the U.S. Food and Drug Administration website http://www.fda.gov/Food/GuidanceRegulation/FederalStateFoodPrograms/ucm2006754.htm





Phyto Kit: Extra bottles, vials, lugol's, rubber gloves, marker



Virginia Estuarine Phytoplankton monitoring

- Chesapeake Bay Monitoring Program (DEQ/ODU)
 - 14 stations



- 7-Chesapeake Bay monthly year-round
- 7-Tidal tributaries monthly March-October
- Full species composition
- Ad hoc bloom sampling
- VDH: Shellfish (DSS&WHC/ODU)



- 69 stations
- Monthly year-round
- Targeted HAB identification
- Targeted toxin screening (based on cell counts)
- CBTOX (VDH:DSS/VIMS)



- 12 stations (2017-2018)
- 4 stations (2019-2020)
- Bi-weekly sampling
- Targeted HAB identification
- Routine toxin analyses

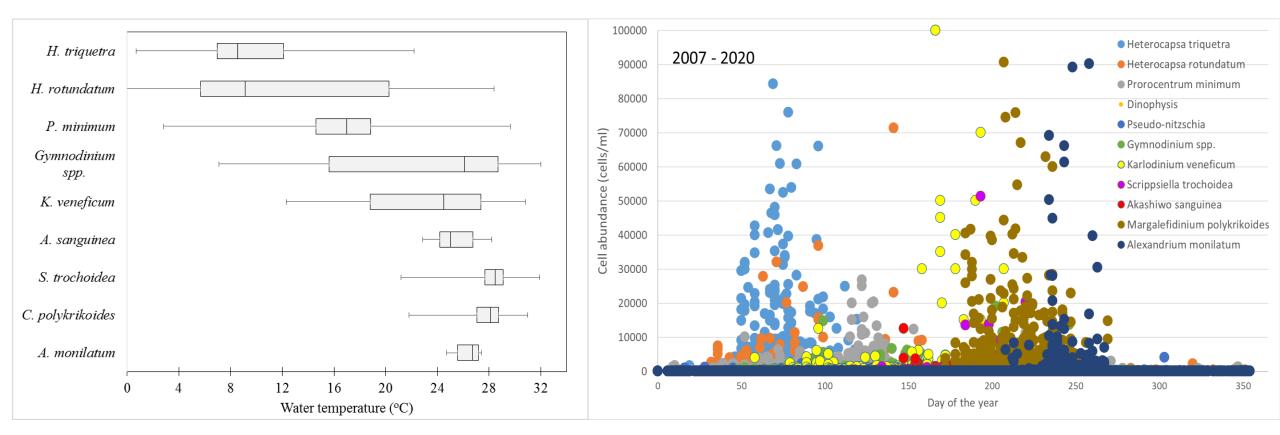


Additional monitoring: ODU, HRSD, ECOHAB Dataflow HRSD (no bloom samples in 2020)

COVID gaps

- No samples from HRSD dataflow from James River so didn't have the data density of previous years
- No CBP sampling March-April; no trib sampling March-June
- No VDH sampling mid-March mid-May
- Did do near-daily sampling in the Elizabeth and Lafayette Rivers throughout the year and were able to document *Heterocapsa* bloom there via IFCB
- Still need to count time series data from Lafayette River

"Normal" bloom progression

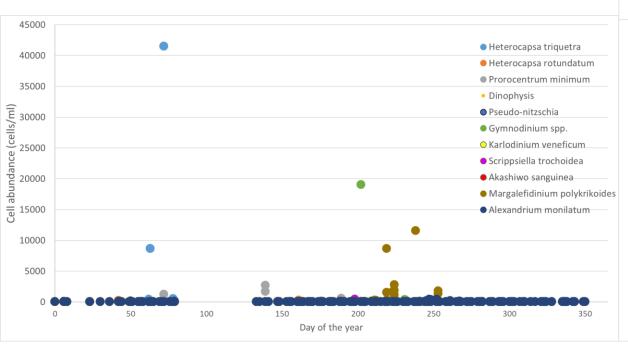


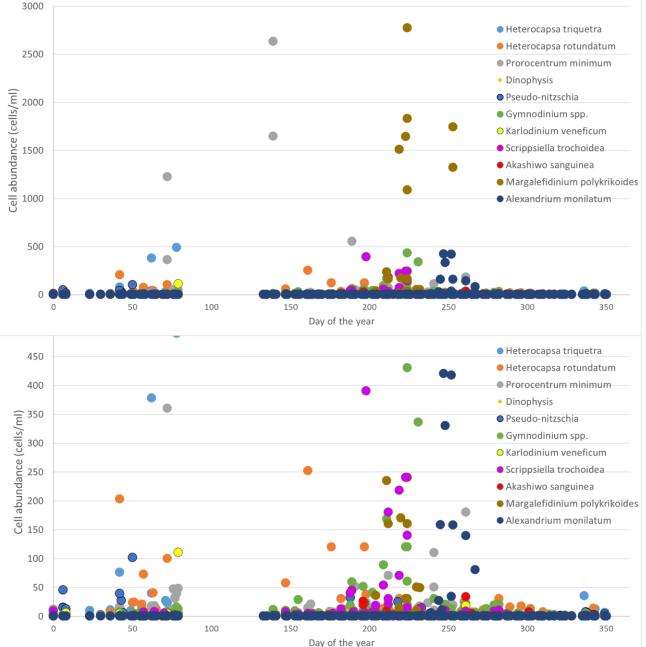
*Pseudo-nitzschia and Dinophysis not abundant enough to make the list

*Removed data where abundances were > 100,000 cells/ml

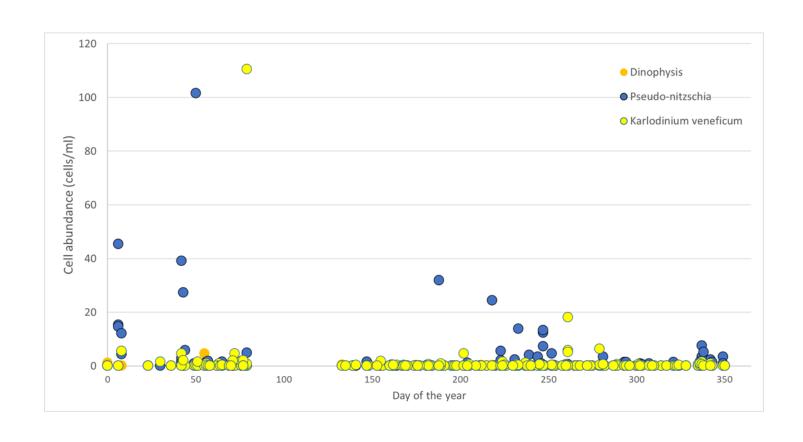
Notable 2020 Blooms:

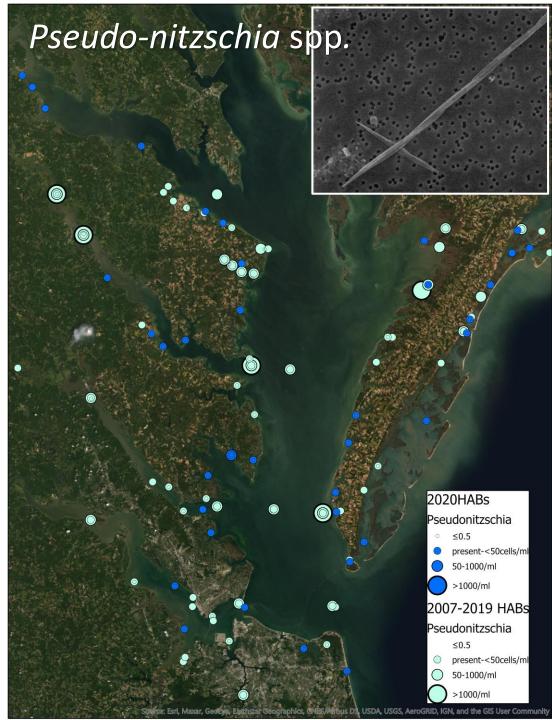
Heterocapsa triquetra (January -March)
Prorocentrum minimum (March – May)
Margalefidinium polykrikoides (July – August)
Alexandrium monilatum (August – September)
But had the full complement of species.



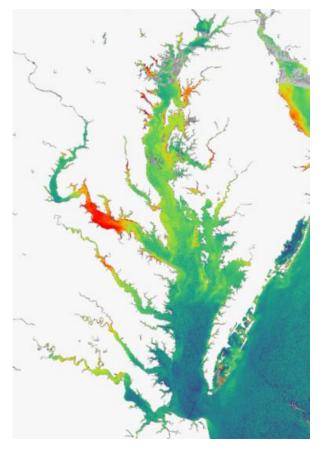


Our less abundant HAB species in 2020





- January 2020 Potomac *Pseudo-nitzschia* event
 - Mixed diatom bloom-Skeletonema dominant

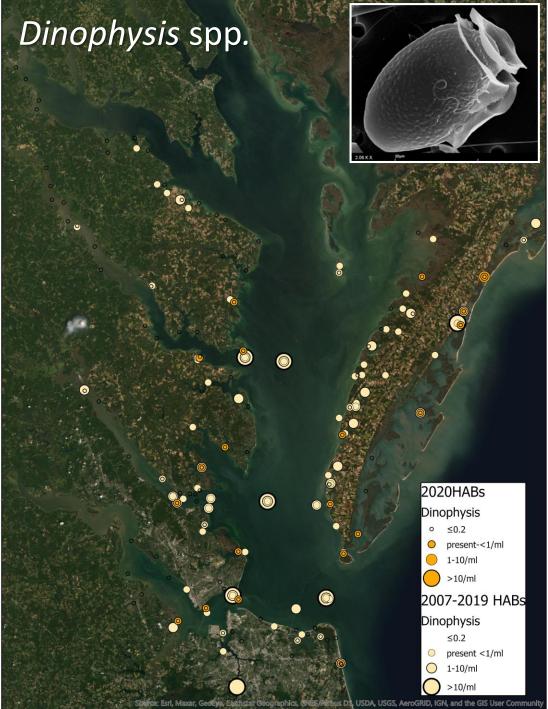


OAA: MODIS-Sentinel imagery-1/1/2020

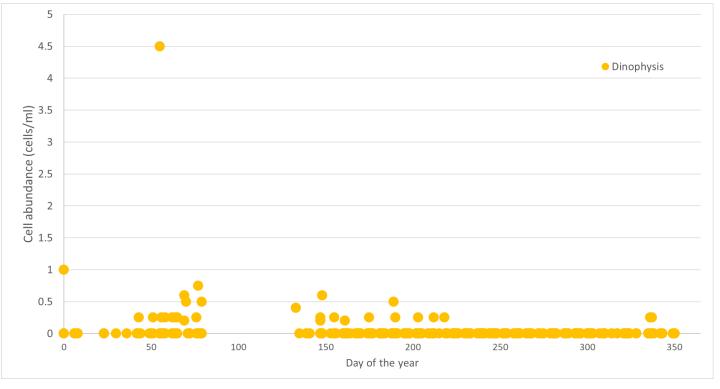


- MD DNR/MDE results ~800- >1000 cells/ml
- Special sample collections
 DSS: January 6, 2020
- Pseudo-nitzschia densities
 52-73 cells/ml
- Toxin BDL

- Generally low cell densities
- Widespread distribution in Chesapeake Bay and seaside Eastern Shore

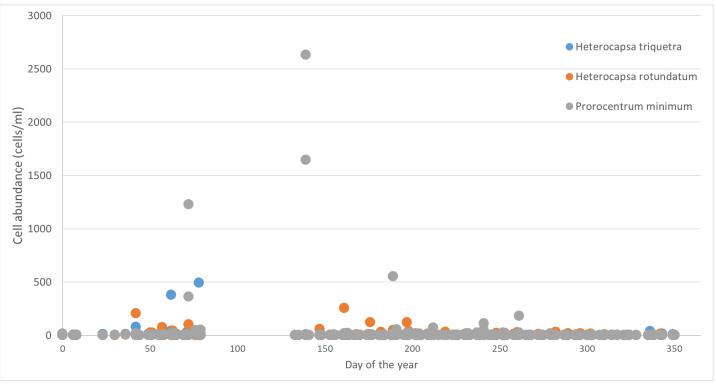


- Generally low cell densities
- Widespread distribution in Chesapeake Bay and seaside Eastern Shore



Heterocapsa triquetra 2020HABs H_triquetra present <500 cells/ml 500-3000 cells/ml >3000 cells/ml 2007-2019 HABs H_triquetra present <500 cells/ml 500-3000 cells/ml

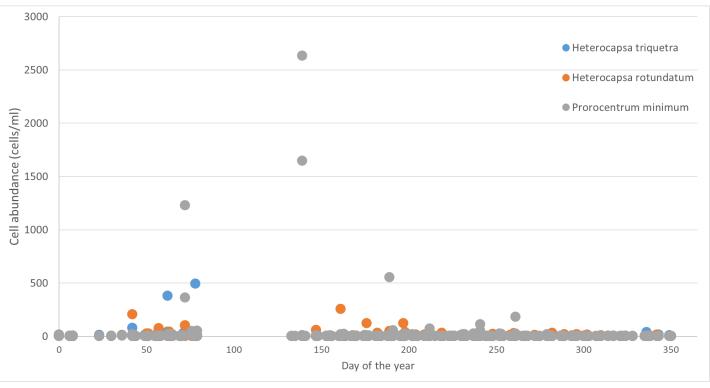
Bloom January - March



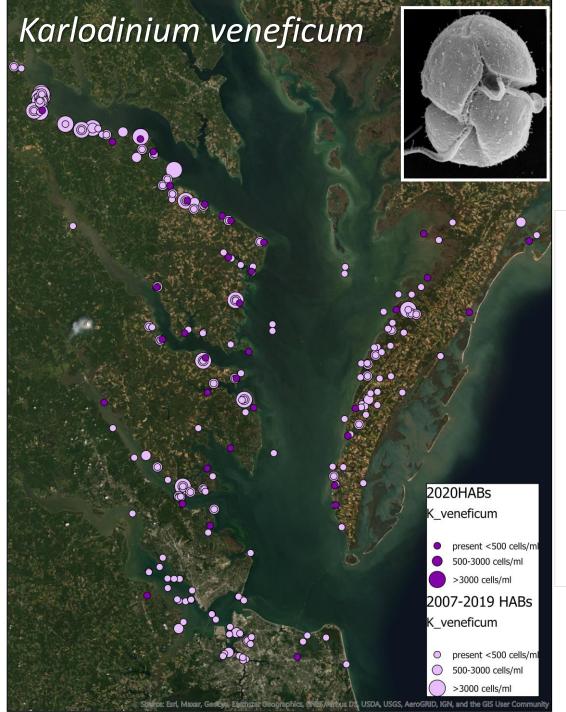
*Removed two *H. triquetra* values > 3000 cells/ml

Prorocentrum minimum 2020HABs P_minimum present <500 cells/m 500-3000 cells/ml >3000 cells/ml 2007-2019 HABs P_minimum present <500 cells/m 500-3000 cells/ml

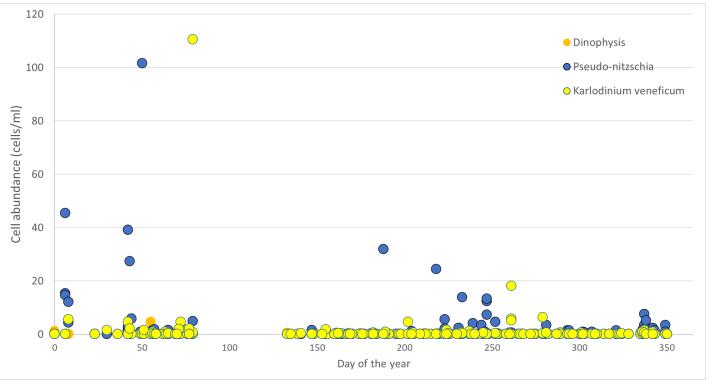
Bloom March – May (under-sampled due to COVID)



*Removed two *H. triquetra* values > 3000 cells/ml



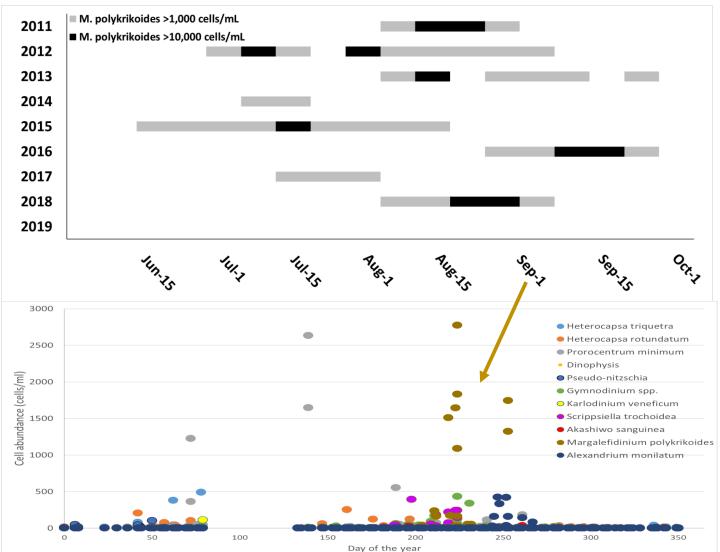
Low abundances but there was a gap in sampling due to COVID



Margalefidinium polykrikoides 2020HABs M. polykrikoides present<1000 cells/ml 1000-10,000 cells/ml >10,000 cells/ml 2007-2019 HABs M. polykrikoides present<1000 cells/ml 1000-10,000 cells/ml

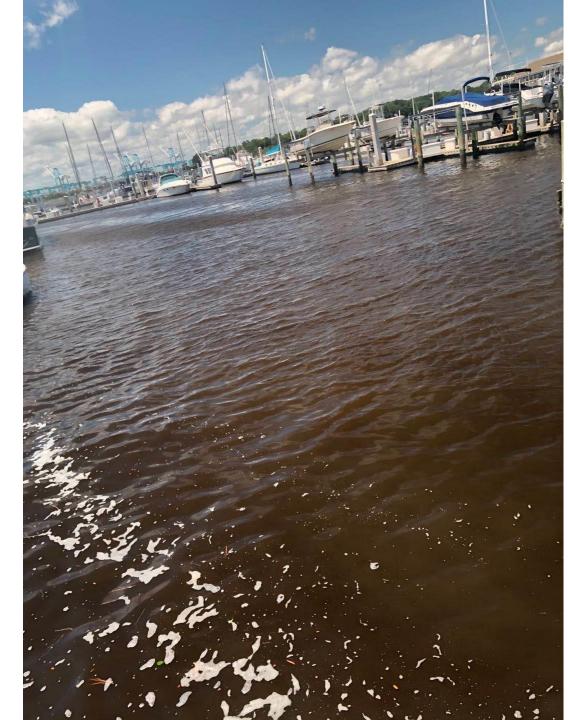
Margalefidinium polykrikoides

- Massive 2020 bloom still counting time series samples
- Initiated in Lafayette River July 20, 2020 & reached oceanfront a week later
- Blooms in most of the last 15 20 years (not in 2019).

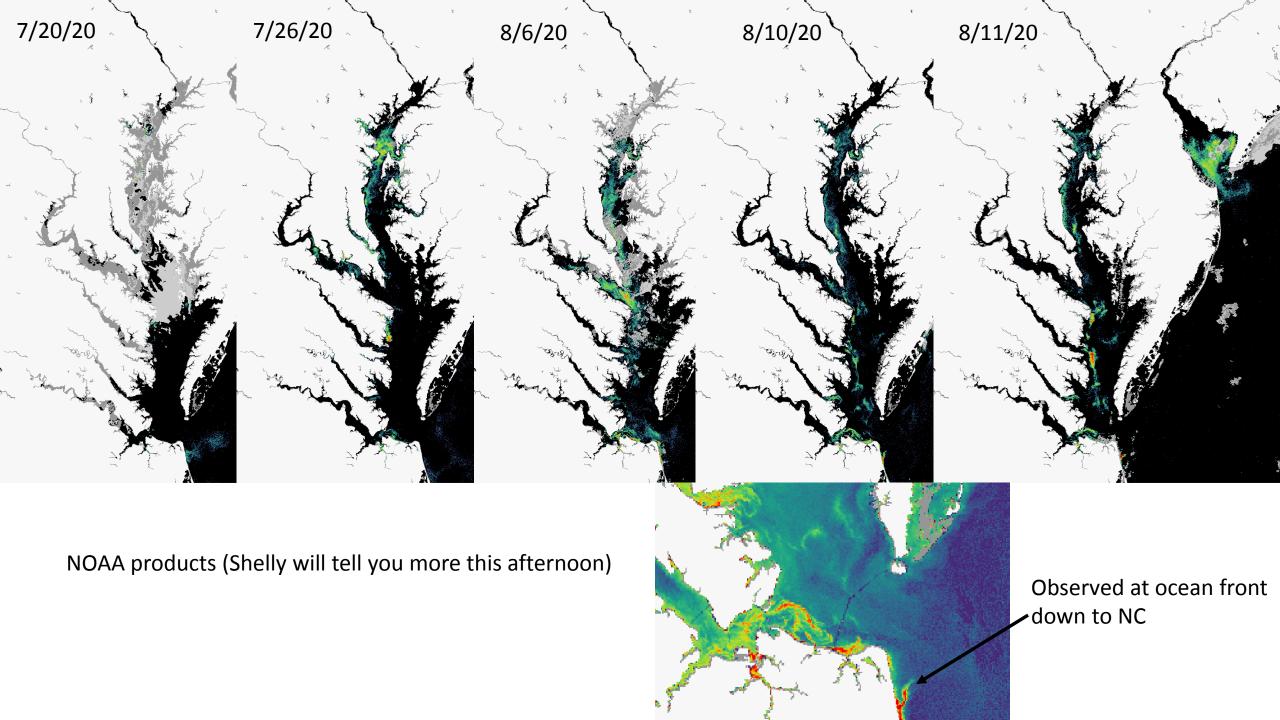


Chlorophyll concentrations in excess of 500 ug/L observed during the day.

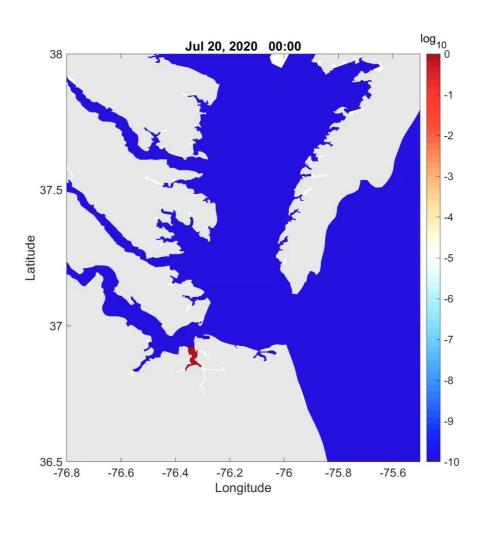


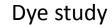






2020 Transport – collaboration with Qin & Shen





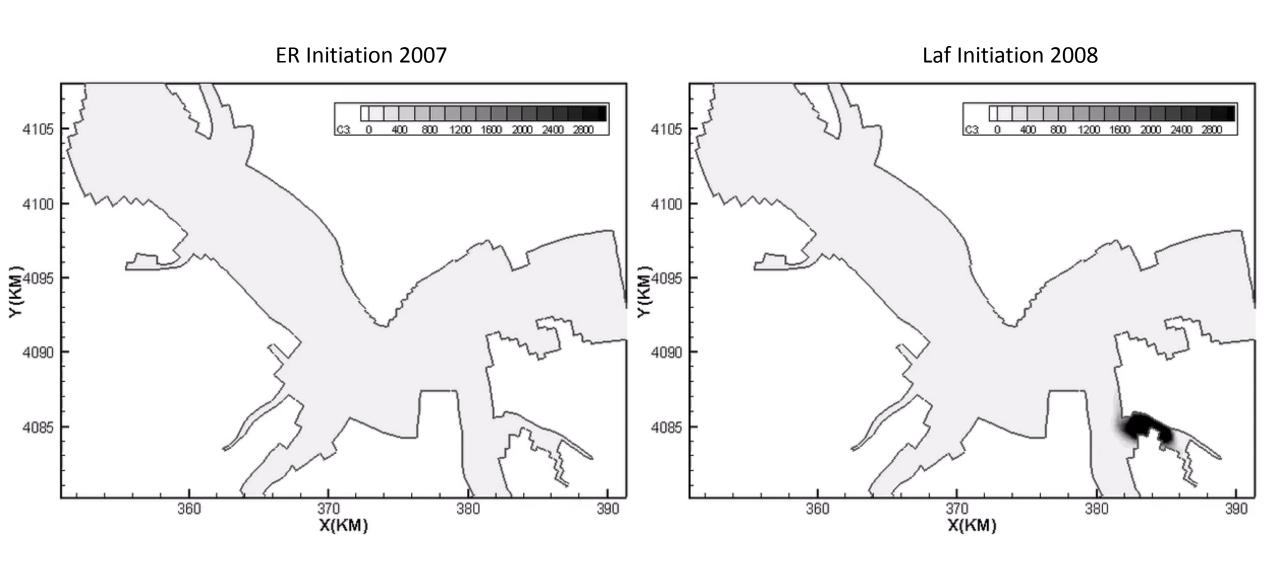
Used calibrated and verified EFDC model for the Ches Bay (Hong & Shen 2012; Du & Shen 2017) and set up model for 2020 using wind data from NOAA, tidal elevation data from 3 station and river discharge data from USGS.

Dye predominantly travelled toward oceanfront, some propagated up Bay.

Big storm on August 4, 2020 which seemed to help transport the bloom.

But bloom persisted.

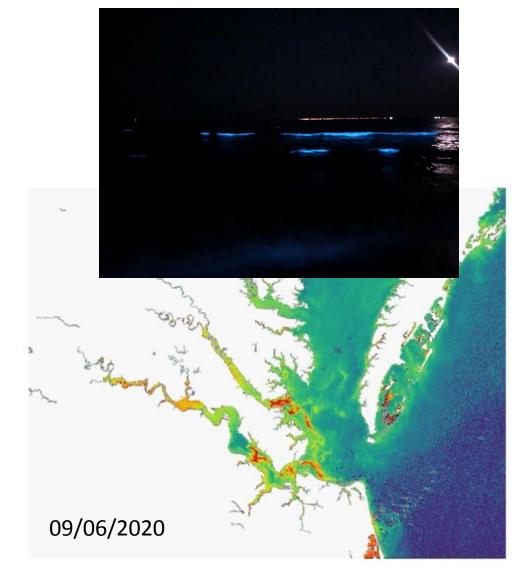
2007 & 2008 Transport

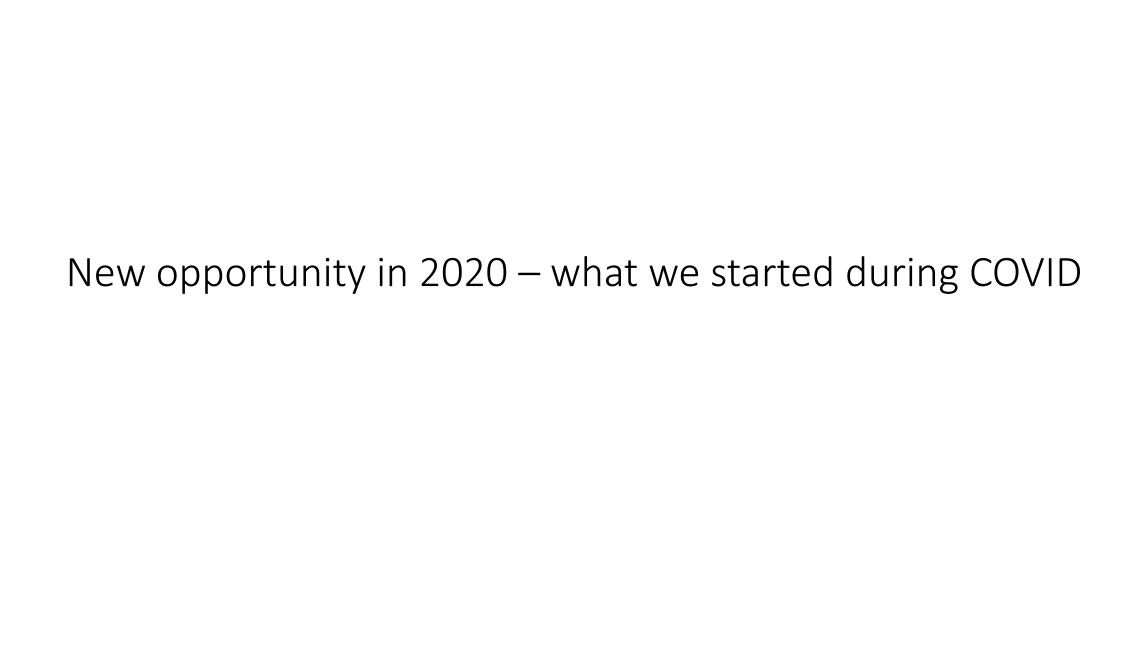


Alexandrium monilatum 2020HABs A monilatum present <1000 cells/ml 1000-10,000 cells/ml >10,000 cells/ml 2007-2019 HABs monilatum present <1000 cells/mL 1000-10,000 cells/mL

Alexandrium monilatum – massive 2020 bloom

- Alexandrium no bloom in 2018 or 2019
- Started in the York River and transported to Ocean View and ocean front

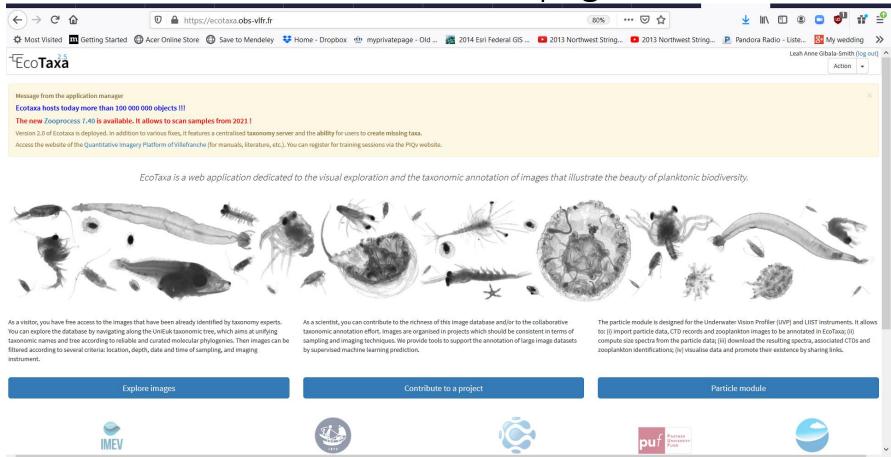




IFCB & EcoTaxa

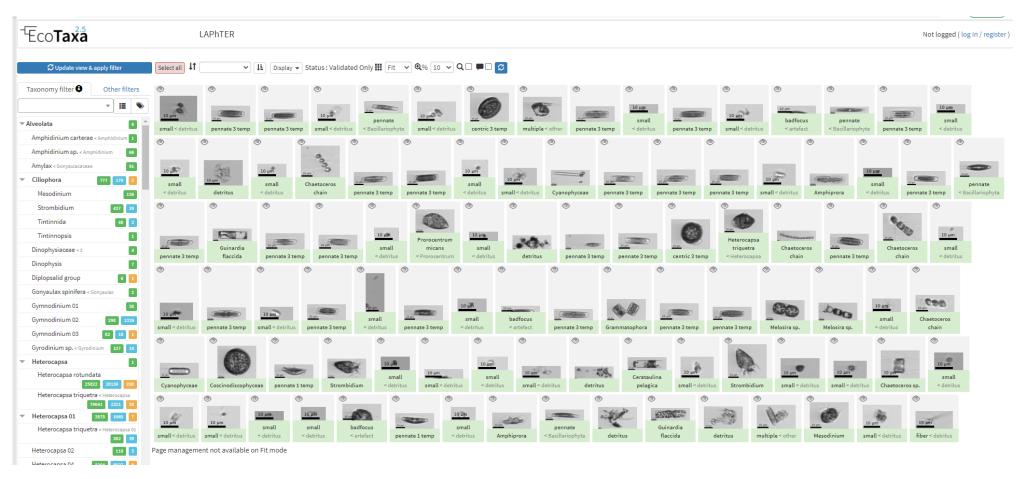
- Imaging flow cytobot (IFCB) images everything, even detritus, in a 5 mL sample (this was just what we fed it)
- Sophie Clayton loads images into EcoTaxa
- Leah and Kathryn (plus 2 undergraduate students) identify taxa and "train" software to identify taxa.

EcoTaxa homepage

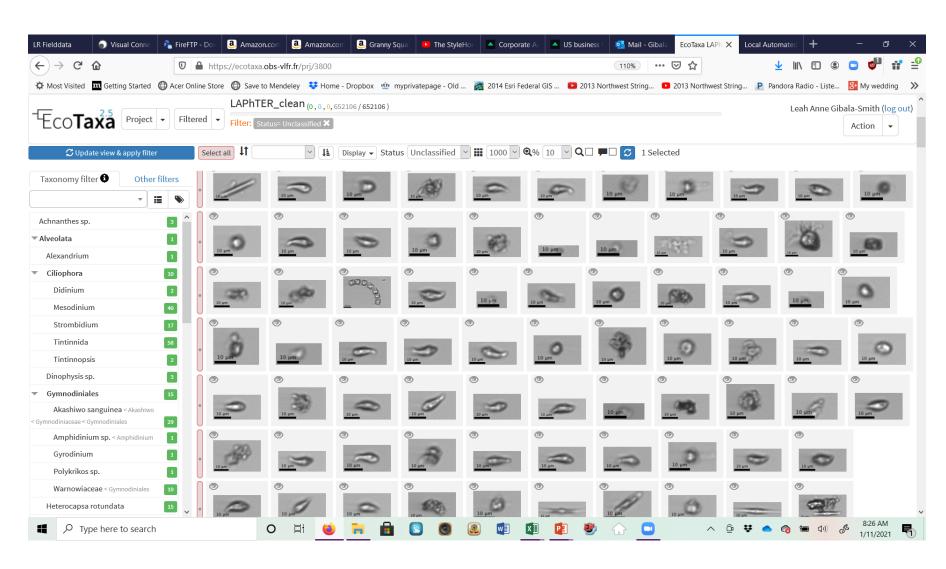


IFCB & EcoTaxa

- Images everything, even detritus, in a 5 mL sample
- We load images into EcoTaxa
- Leah and Kathryn (plus 2 undergraduate students) identify taxa and "train" software to identify taxa.



This is an unclassified upload of images from IFCB into ECOTAXA project ready for sorting and validating



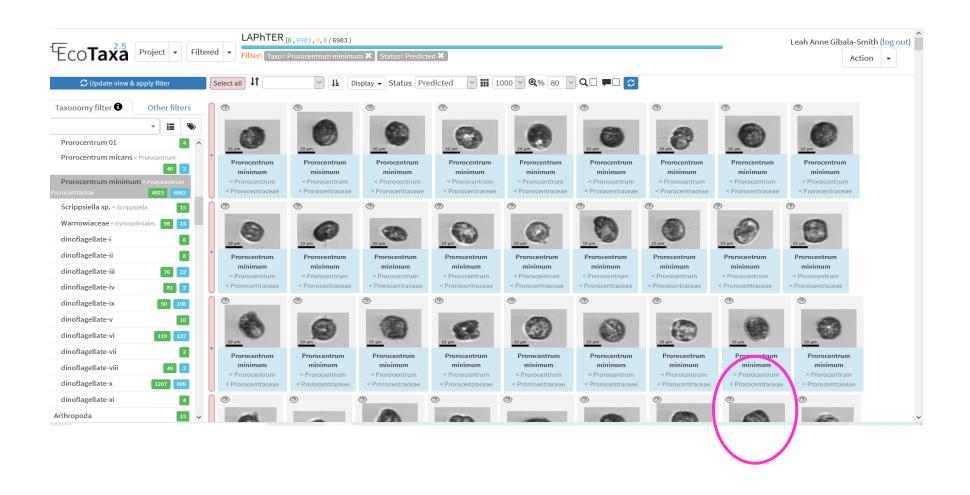
Steps in procedure

- 1. Upload images and metadata from IFCB into EcoTaxa
- 2. Manually classify organisms requires creation of new categories within taxonomic tree
- 3. Run predictive model on unclassified images to group them into taxa categories
- 4. Clean predictions by manually validating a portion of the correctly sorted cells and and moving incorrectly sorted cells into their correct taxonomic group
- 5. Run prediction on unvalidated sorted cells and manually validate/correct a portion of them
- 6. REPEAT

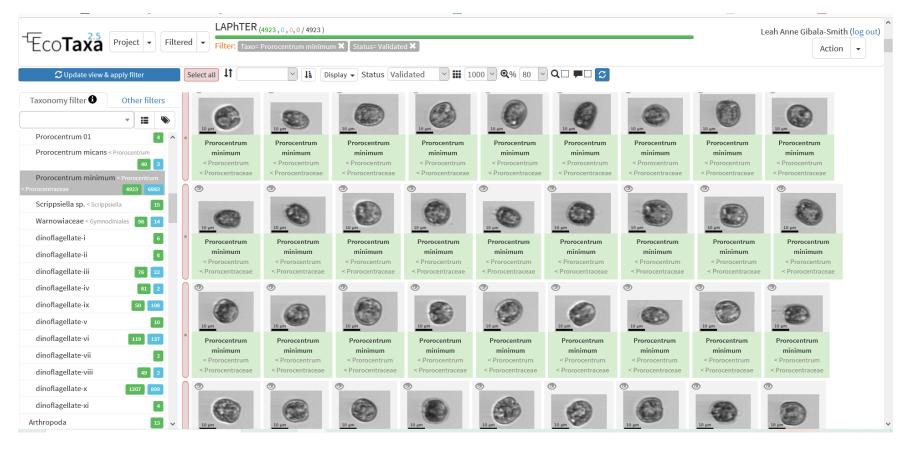


Garbage vs cells!!!

Prediction run – Only 1 cell predicted is a *P. minimum*



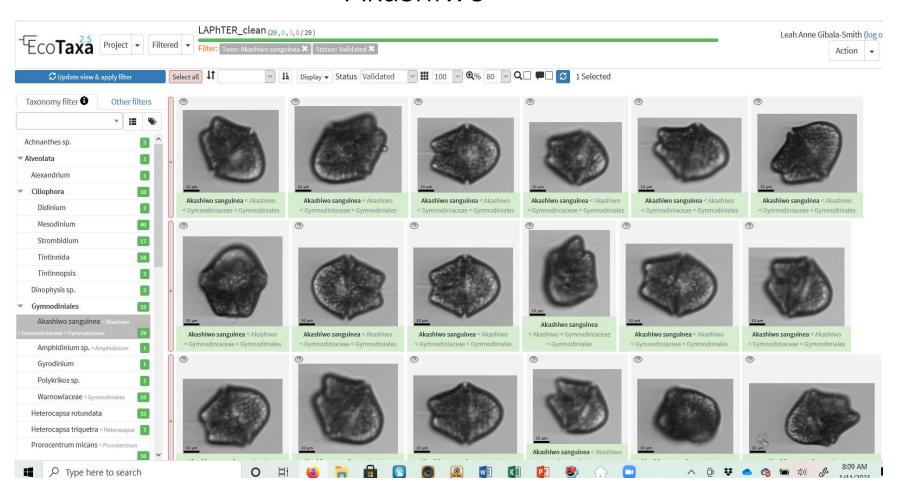
Validated and corrected



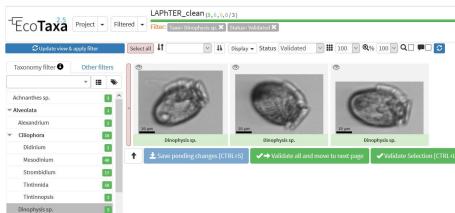
There are inconsistencies in model predictions for some organisms because cell morphology can be highly variable So, make sub-categories to account for life stages, etc. - more on this later – and rerun the model, it is an iterative process

Eventually, we can group cells and train the instrument to count them in individual samples

Akashiwo

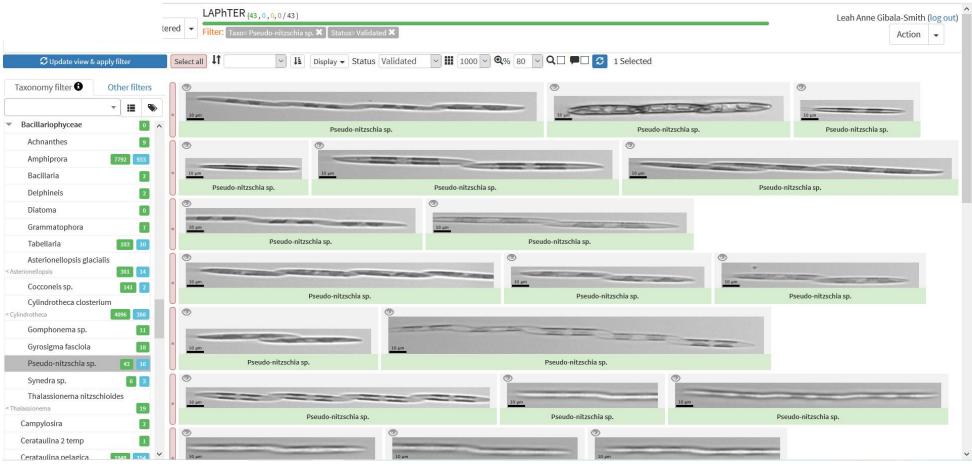


Dinophysis

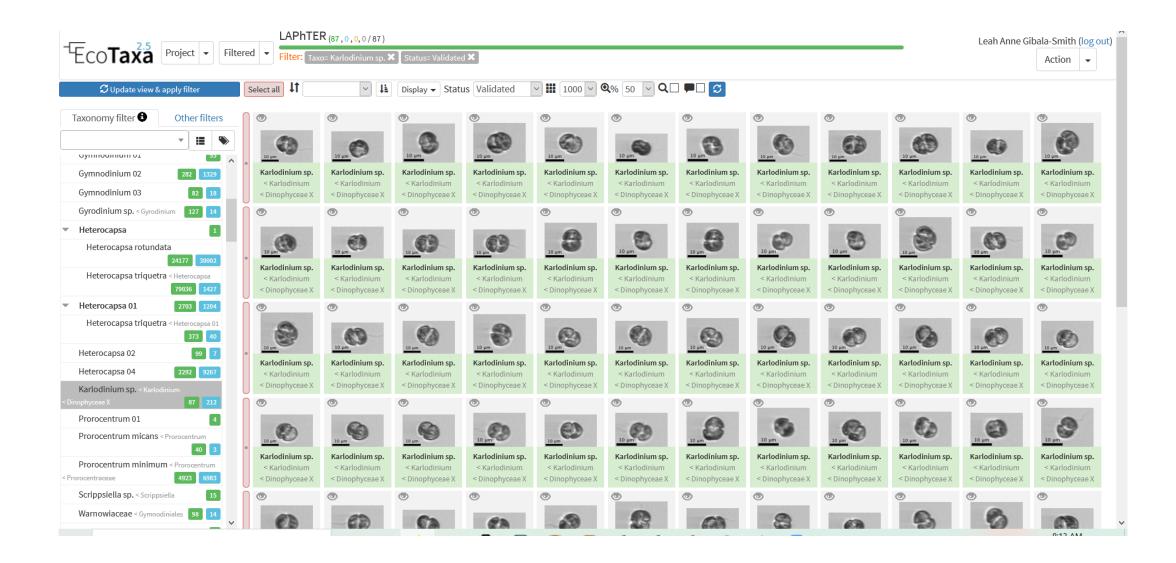


Even the rare ones! We can examine more sample in detail so this may help us!

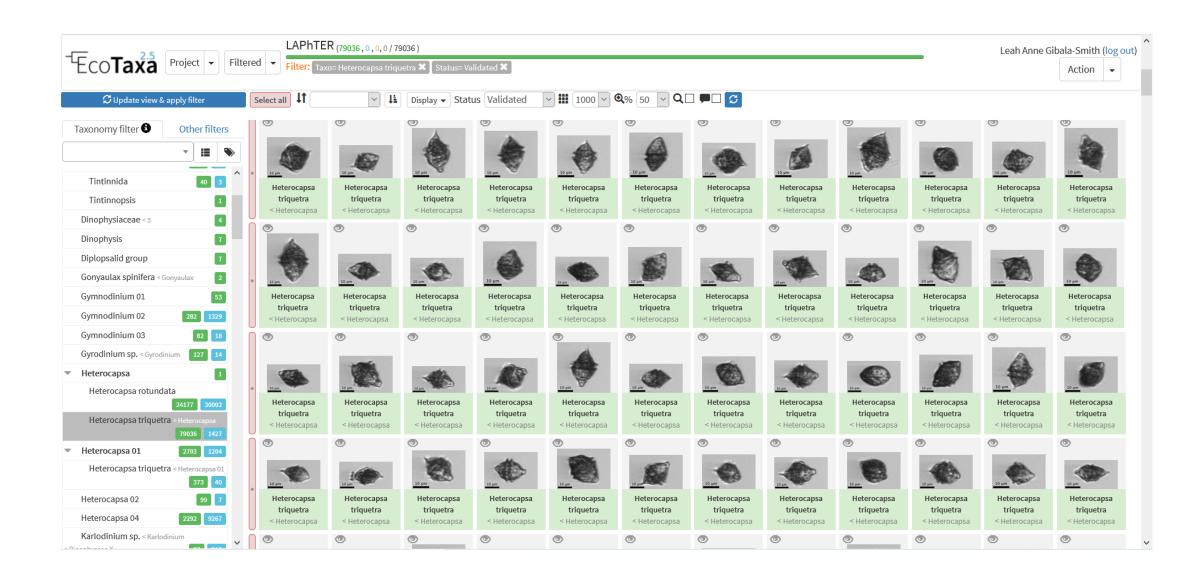
Pseudo-nitzschia



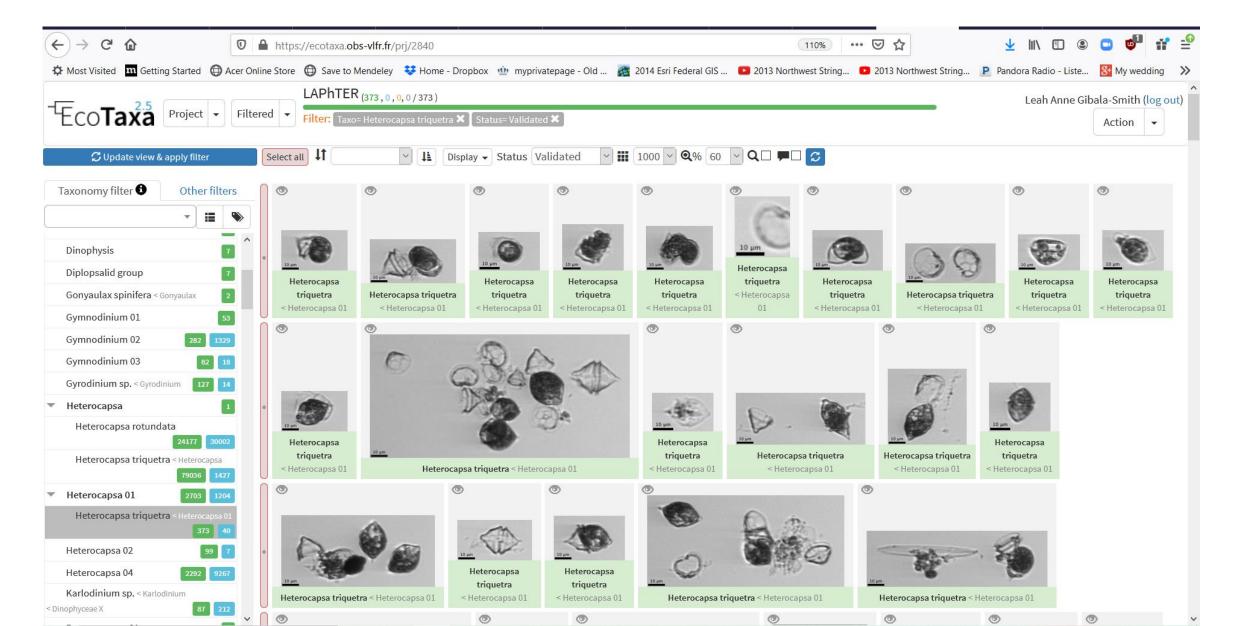
Karlodinium



Heterocapsa triquetra – we're also learning about the organisms themselves!



Heterocapsa triquetra – we're seeing different life stages

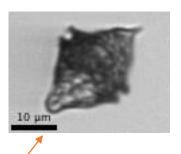


Heterocapsa triquetra Peridinale

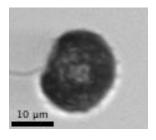
Asexual reproduction observed: Eleutheroschisis: theca sheds before or after cell division Sexual reproduction not observed Heterocapsa groupings:

Heterocapsa 01

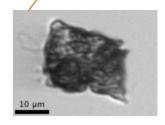
stress



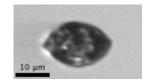
Heterocapsa triquetra Vegetative cell



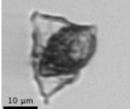
Heterocapa 04 antapical view of the vegetative cell



Heterocapsa 02Binary fission

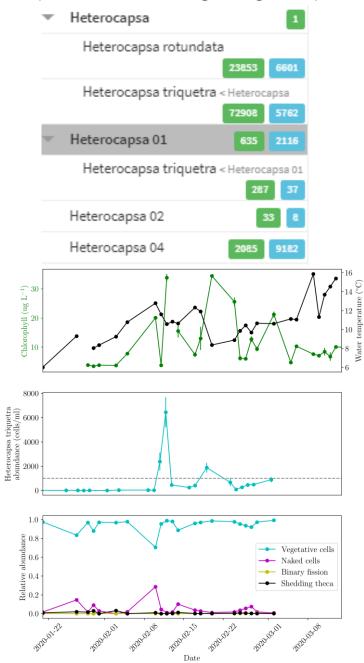


Ecdysis has occurred -"naked" cells that have shed their theca (part of asexual reproduction) OR can be the result of

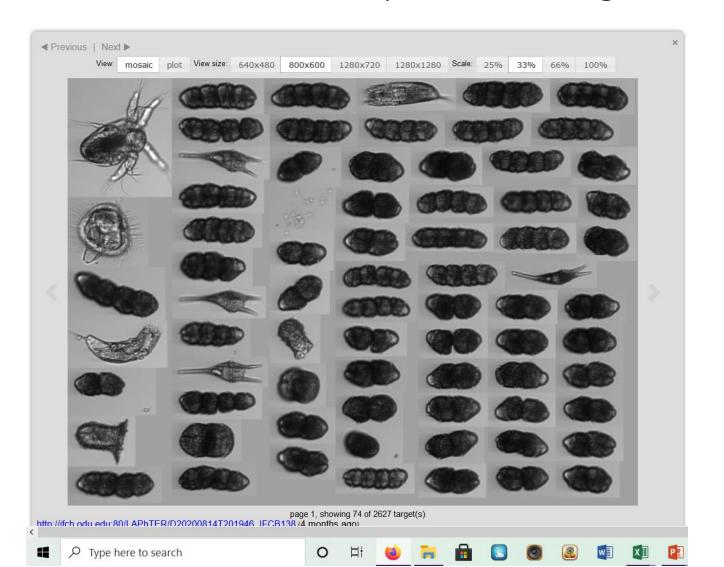


Heterocapsa triquetra < Heterocapsa 01 Cells in the process of shedding their theca but still have an intact pellicle

ECOTAXA CATEGORY TREE & definitions (intermediate working arrangement)



IFCB live dashboard – *Margalefidinium* bloom – images not uploaded to Ecotaxa yet We know life stages and chain length may play an important role in the initiation and persistence of these blooms and plan to interrogate the database.



2021 program plans

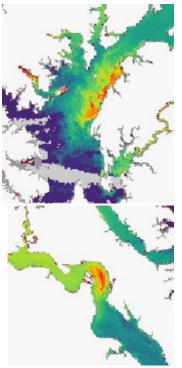
- Continue VDH & CBP sampling and merging of databases
- Resume HRSD dataflow sampling after COVID hiatus
- Continue sampling at the Lafayette River time series
- Merging of databases
- Continue IFCB training
 - Also getting Flow Cam loaner this winter
- Building off multiple programs VDH, CBP, HRSD, ECOHAB
- Continued discussions with partners
 - Phytoplankton methods- IFCB, remote sensing
 - Management strategies

New year's resolutions/goals

- Find a way to get a dedicated IFCB
- Year-round sampling and analyses with undergrads and instruments
- Laboratory experiments
 - Isolation of cultures
 - Production of toxins
 - Life cycle events that influence blooms
- More student involvement
- New funding to tie in research with the monitoring to better advise management







Thank you!

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VIRGINIA DEPARTMENT OF HEALTH To protect the health and promote the well-being of all people in Virginia







People:

Leah Gibala-Smith Kathryn Mogatas **Todd Egerton** Michael Echevarria Eduardo Perez Vega Alfonso Macias Tapia Yifan Zhu Peter Bernhardt Sophie Clayton Shelly Tomlinson Qubin Qin Jian Shen All the field crew at DEQ, VDH, & HRSD

